

Mouse Glutamine Synthetase (GS) ELISA Kit

Cat. No.: EJ1513254 | Pack size: 48T / 96T | Storage: Store at 2-8 °C

Overview

Glutamine synthetase (GS) plays a crucial role in nitrogen metabolism by catalyzing the condensation of glutamate and ammonia to form glutamine.

This kit uses a double-antibody sandwich ELISA to detect mouse glutamine synthetase (GS). The color intensity at 450 nm is positively correlated with the concentration of mouse GS in the sample.

Key Features

- Double-antibody sandwich ELISA for mouse GS
- Detection range: 0.31 ng/mL - 20 ng/mL
- Sample types include cell culture supernatants, cell lysate, cerebrospinal fluid, plasma, serum and tissue homogenates
- Intra- and inter-assay CV values are both <10%
- Detection duration is approximately 3.5 hours, excluding equilibration and sample preparation

Product Information

Item	Description
Synonyms	GLNS GLUL Glutamate-Ammonia Ligase Glutamate decarboxylase
Specifications & Purity	BioReagent, for enzyme immunoassay (ELISA)
Application	ELISA
Sample Type	Cell culture supernatants, cell lysate, cerebrospinal fluid, plasma, serum, tissue homogenates
Detection Range	0.31 ng/mL - 20 ng/mL
Method of Detection	ELISA; sandwich method with two antibodies
Detected Species Object	Mouse
Specificity	Detects mouse GS with no significant cross-reactivity to other homologs.

Item	Description
Detection Instrument / Wavelength	Microplate reader / 450 nm
Repeatability	Intra- and inter-assay CV <10%
Storage & Shipping	Store at 2-8 °C; shipped on wet ice; shelf life 6 months

Contents & Storage

Cat. No.	Component	48T	96T	Storage
EJ1513254A	Pre-coated Assay Plate	48 wells	96 wells	2-8 °C
EJ1513254B	Standard	1 vial	2 vials	2-8 °C
EJ1513254C	Universal Diluent	1 x 20 mL	2 x 20 mL	2-8 °C
EJ1513254D	Biotin-antibody (100x)	60 µL	120 µL	2-8 °C
EJ1513254E	Streptavidin-HRP (100x)	60 µL	120 µL	2-8 °C
EJ1513254F	Wash Buffer (20x)	1 x 10 mL	2 x 10 mL	2-8 °C
EJ1513254G	TMB Substrate	5 mL	10 mL	2-8 °C
EJ1513254H	Stop Solution	3 mL	6 mL	2-8 °C
EJ1513254I	Plate Sealer	4 sheets	4 sheets	2-8 °C

Note: EJ1513254B, EJ1513254D, EJ1513254E and EJ1513254F shall be diluted according to the instructions.

Shelf life: Each component has a shelf life of 6 months under corresponding storage conditions.

Materials Required But Not Supplied

Item	Recommended Specification	Purpose
Microplate reader	450 nm	Measurement of OD value
High-precision pipettes and tips	0.5-10 µL, 5-50 µL, 20-200 µL, 200-1000 µL	Sample and reagent dispensing

Item	Recommended Specification	Purpose
37 °C incubator	Stable temperature control	Plate incubation
Distilled or deionized water	Freshly prepared	Preparation of 1x wash buffer
Microcentrifuge tubes	Clean, labeled EP tubes	Standard and sample dilution
Wash bottle or plate washer	Compatible with 96-well plates	Plate washing

Experimental Principle

This kit adopts a double-antibody sandwich enzyme-linked immunosorbent assay (ELISA). Samples, standards, biotin-labeled detection antibody and HRP conjugate are sequentially added to microwells pre-coated with mouse GS capture antibody.

After incubation and washing, TMB substrate is used for color development. TMB is catalyzed by HRP to blue and converted to yellow under acidic conditions. The color intensity is positively correlated with the concentration of mouse GS in the sample. The absorbance (OD value) is measured at 450 nm and the sample concentration is calculated from the standard curve.

Sample Handling and Requirements

1. The kit detection range does not necessarily equal the analyte concentration range in samples. Estimate sample concentration from literature or a preliminary experiment; dilute or concentrate samples appropriately if needed.
2. If the sample type is not listed in this manual, run a preliminary experiment to verify detection validity.
3. Serum: allow whole blood collected in serum separation tubes to stand at room temperature for 2 hours or overnight at 2-8 °C, then centrifuge at 1000 x g for 20 minutes and collect the supernatant. Store at -20 °C or -80 °C if needed; avoid repeated freeze-thaw cycles.
4. Plasma: collect using EDTA or heparin as anticoagulant, centrifuge at 1000 x g for 15 minutes at 2-8 °C within 30 minutes of collection, and use the supernatant for testing. Store at -20 °C or -80 °C if needed; avoid repeated freeze-thaw cycles.
5. Tissue homogenate: rinse tissue with pre-chilled PBS (0.01 M, pH 7.4) to remove residual blood. Weigh and mince tissue, add PBS at about 1:9 (w/v), homogenize on ice, then further lyse by sonication or repeated freeze-thaw if needed. Centrifuge at 5000 x g for 5-10 minutes and use the supernatant.

6. Cell culture supernatant: centrifuge at 1000 x g for 20 minutes and use the supernatant for testing. Store at -20 °C or -80 °C if needed; avoid repeated freeze-thaw cycles.
7. Cell lysate: wash cells with pre-chilled PBS, collect by centrifugation at 1000 x g for 5 minutes, wash three times with PBS, resuspend in 150-200 µL PBS per 1×10^6 cells, lyse by freeze-thaw or sonication, then centrifuge at 1500 x g for 10 minutes at 2-8 °C and use the supernatant.
8. Other biological samples: centrifuge at 1000 x g for 20 minutes and use the supernatant for testing.
9. Samples should be clear and transparent. Remove suspended matter by centrifugation. Hemolyzed samples should not be used because hemolysis interferes with results.
10. If testing within 1 week, store samples at 4 °C. For delayed testing, aliquot and store at -20 °C for up to 1 month or -80 °C for up to 6 months; avoid repeated freeze-thaw cycles.

Sample Dilution Protocol

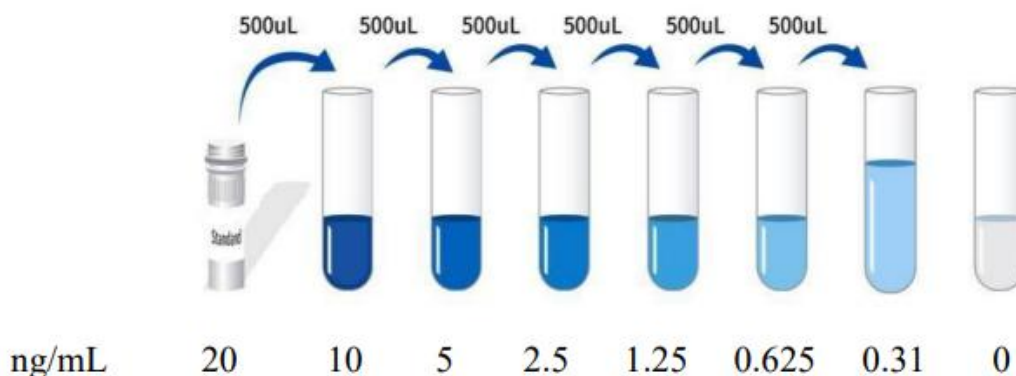
Estimate the sample concentration range in advance. If dilution is required, follow the protocol below. In each step, use at least 3 µL for pipetting and do not exceed 100-fold dilution per step. Mix thoroughly after each dilution and avoid bubbles.

1. 100-fold dilution: add 5 µL sample to 495 µL general diluent.
2. 1000-fold dilution: add 5 µL sample to 95 µL general diluent (20-fold), then add 5 µL of the 20-fold dilution to 245 µL general diluent (50-fold); total dilution = 1000-fold.
3. 100,000-fold dilution: add 5 µL sample to 195 µL general diluent (40-fold), then add 5 µL of the 40-fold dilution to 245 µL general diluent (50-fold), then add 5 µL of the 2000-fold dilution to 245 µL general diluent (50-fold); total dilution = 100,000-fold.

Pre-Test Preparation

1. Remove the kit from the refrigerator 10 minutes in advance and equilibrate it to room temperature.
2. Prepare standard gradient working solution: add 1 mL universal diluent to the lyophilized standard, stand for 15 minutes until completely dissolved, and mix gently. The top standard concentration is 20 ng/mL. Prepare serial dilutions to 20, 10, 5, 2.5, 1.25, 0.625, 0.31, 0 ng/mL.

Serial dilution method: place 7 EP tubes, each containing 500 µL universal diluent. Transfer 500 µL of the top standard working solution to the first tube and mix; repeat sequential transfer and mixing. The last tube is used as the blank well and no liquid is transferred from the penultimate tube.



3. Prepare biotin-antibody working solution: centrifuge the 100x concentrated biotin-antibody at 1000 x g for 1 minute 15 minutes before use. Dilute to 1x with universal diluent (for example, 10 μ L concentrate + 990 μ L diluent). Prepare freshly before use.
4. Prepare enzyme conjugate working solution: centrifuge the 100x concentrated enzyme conjugate at 1000 x g for 1 minute 15 minutes before use. Dilute to 1x with general diluent. Prepare freshly before use.
5. Prepare 1x washing solution: add 10 mL of 20x washing solution to 190 mL distilled water. If crystals are present in concentrated wash buffer, allow it to reach room temperature and dissolve completely before preparation.

Procedure

1. Remove the required strips from the aluminum foil bag after equilibration at room temperature for 10 minutes. Seal the remaining strips in a resealable bag and return to 4 °C.
2. Sample addition: add 100 μ L of samples or standards of different concentrations to the corresponding wells. Add 100 μ L universal diluent to blank wells. Cover with a plate sealer and incubate at 37 °C for 60 minutes. Duplicate wells are recommended for all samples and standards.
3. Add biotinylated detection antibody: discard liquid without washing, then add 100 μ L biotinylated detection antibody working solution to each well. Cover and incubate at 37 °C for 60 minutes.
4. Wash plate: discard liquid, add 300 μ L 1x washing buffer to each well, stand for 1 minute, discard wash buffer and pat dry on absorbent paper. Repeat washing 3 times or use a plate washer.
5. Add enzyme conjugate working solution: add 100 μ L to each well. Cover and incubate at 37 °C for 30 minutes.
6. Wash plate 5 times as described above.
7. Add substrate: add 90 μ L TMB substrate to each well. Cover and incubate at 37 °C in the dark for 15 minutes.

8. Add stop solution: add 50 μ L stop solution to each well and immediately measure OD at 450 nm.

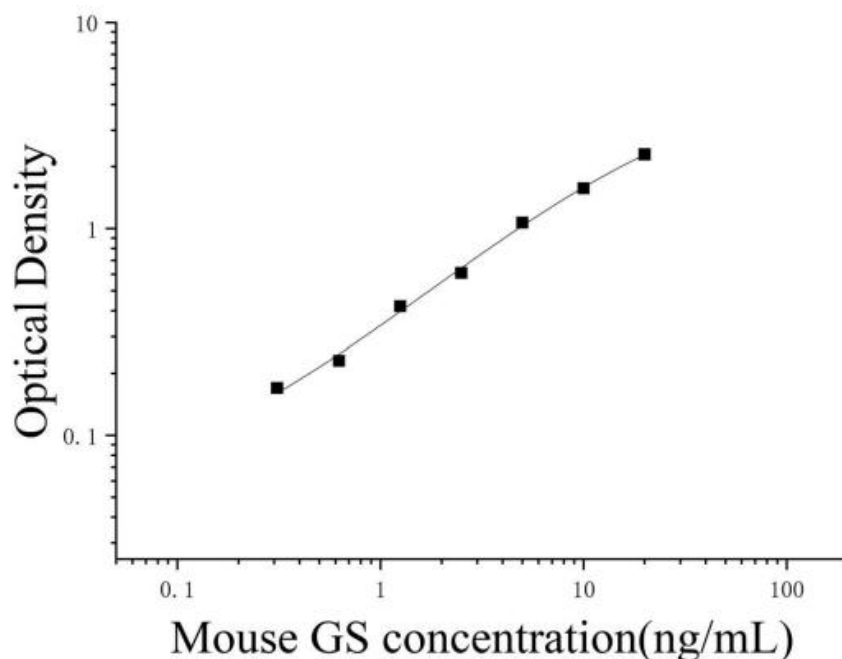
Calculation of Experimental Results

1. Calculate the average OD value of duplicate standards and samples, then subtract the blank OD value to obtain the corrected OD value. Plot a standard curve using a four-parameter logistic function, with concentration on the x-axis and OD value on the y-axis.
2. If the sample OD value exceeds the upper limit of the standard curve, dilute the sample appropriately and re-test. Multiply the final concentration by the dilution factor.

Typical Data and Reference Curve

The following data and curve are for reference only. Users must establish a standard curve based on each experiment.

Concentration (ng/mL)	20	10	5	2.5	1.25	0.625	0.31	0
OD value	2.39	1.67	1.17	0.71	0.52	0.33	0.27	0.1
Corrected OD value	2.29	1.57	1.07	0.61	0.42	0.23	0.17	-



Kit Performance

1. Repeatability: Intra-assay CV < 10%; inter-assay CV < 10%.
2. Recovery: Add three different concentrations of mouse GS to healthy mouse serum, plasma, and cell lysate, then calculate recovery.

Sample Type	Range (%)	Average Recovery (%)
Serum (n=8)	84-101	96
Plasma (n=8)	92-105	102
Cell lysate (n=8)	96-108	105

3. Linearity of dilution: Add high-concentration mouse GS to four healthy mouse serum, plasma, and cell lysate samples, then dilute within the dynamic range of the standard curve to evaluate linearity.

Dilution Ratio	Recovery (%)	Serum	Plasma	Cell lysate
1: 2	Range	84-95	88-96	90-110
1: 2	Average recovery	91	93	96
1: 4	Range	89-103	87-108	105-115
1: 4	Average recovery	94	98	109

Safety & Precautions

1. Bring all reagents to room temperature (20-25 °C) before use and store reagents at 2-8 °C immediately after use.
2. Strictly follow the specified incubation time and temperature. Inadequate incubation or washing may cause inaccurate results.
3. Do not allow microwells to dry during the assay. Drain wells thoroughly before adding substrate.
4. Clean residual liquid and fingerprints from the well bottoms before reading optical density.
5. Use colorless TMB substrate only. Do not use substrate that has turned blue before use.
6. Avoid cross-contamination between reagents and samples. Do not mix kit components from different catalog numbers or lots.
7. Protect reagents and plates from strong light during storage and incubation.

8. Handle all biological samples according to established laboratory biosafety procedures.

Quality Control

QC Item	Method	Acceptable Range
Standard curve	Run standards in duplicate	Curve covers specified detection range and blank-corrected OD values are suitable for interpolation.
Precision	Duplicate measurements	Intra- and inter-assay CV <10%.
Kit integrity	Receipt and storage inspection	Components complete; stored at 2-8 °C; expiry date and lot number legible.

Troubleshooting

Issue	Possible Causes	Corrective Action
Poor standard curve linearity	Incorrect standard dilution	Dissolve and serially dilute standards exactly as described. Mix thoroughly and avoid bubbles.
Poor standard curve linearity	Inaccurate pipetting	Calibrate pipettes regularly and check tip sealing. Run standards in duplicate.
Poor standard curve linearity	Evaporation of reaction solution	Seal the plate with film during incubation.
Poor standard curve linearity	Inadequate washing	Use sufficient wash buffer volume and repeat the recommended wash times.
Weak or no color development	Insufficient incubation time or incorrect temperature	Ensure full incubation at 37 °C for each specified step.
Weak or no color development	Incorrect reagent dilution	Verify dilution of biotin-antibody, HRP conjugate and wash buffer.
Weak or no color development	Inactivated enzyme conjugate	Check for color development with conjugate and substrate; replace reagents if needed.
Low OD values	Incorrect reader wavelength or delayed reading	Read at 450 nm immediately after adding stop solution.

Issue	Possible Causes	Corrective Action
OD value outside standard range	Sample concentration too high or too low	Dilute or concentrate the sample based on a preliminary experiment and re-test.
High background	Contaminated substrate or excessive development time	Replace substrate if needed and control color development time.
High background	Inadequate washing	Wash thoroughly and pat dry on absorbent paper after each wash sequence.

Recommended Applications

ELISA detection of mouse GS · Serum samples · Plasma samples · Tissue homogenates · Cell-culture supernatants · Cell lysates · Cerebrospinal fluid · Double-antibody sandwich immunoassay research

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- This product is not a CE-marked in vitro diagnostic device under IVDR (EU) 2017/746 and is not an FDA-cleared device under 21 CFR. Use is restricted to verified businesses, institutions, and qualified professionals for research and development purposes.

- Where any kit component is classified as hazardous under CLP (EC) 1272/2008 or OSHA HCS (29 CFR 1910.1200), the product Safety Data Sheet (SDS) takes precedence over this document for handling, storage, and disposal information.
- Performance depends on sample type, sample condition, handling, and operator technique. Users are responsible for validating the kit for their specific application.
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